

© by Springer-Verlag 1980

# Cytoplasmic Male Sterility in Barley

Part 7: Nuclear Genes for Restoration

### H. Ahokas

Department of Genetics, University of Helsinki, Helsinki (Finland)

Summary. Restoration in the *msm1* cytoplasm of barley (Hordeum vulgare L. s.l.) was studied from the standpoint of population biology and physiological effects on kernel protein. Restorer genes of 82 accessions of wild barley (ssp. spontaneum) from Israel were determined. 38% of the accessions were maintainers of sterility, 48% were partial restorers, and 14% were restorers. Fourteen dominant restorer genes are described, and evidence for three cases of allelism to Rfm1a is presented. The restorer accessions and their designated gene symbols are: PI 282636 (Rfm,,e), PI 282637 (Rfm,,f), PI 282646 (Rfm,,g), PI 284742 (Rfm,,h), PI 284743 (Rfm,,i), PI 284753 (Rfm,,j), PI 284755 (Rfm1d), PI 296838 (Rfm,,k), PI 296850 isolate 16/7 (Rfm,,l), PI 296853 (Rfm,,m), PI 296856 (Rfm1b), PI 296899 (Rfm,,n), PI 296919 (Rfm1c), PI 296944 (Rfm, o). PI 296850 was found to contain both a restorer and a non-restorer genotype. None of the PI accessions with a restorer gene is a carrier of an msm1-type male sterilizing cytoplasm. In the present sample, plants with restoration ability occurred with a higher frequency in the material from the Judean Foothills than that from the other regions of Israel. The greater adaptive value of plants with restoration ability on certain soil associations in semiarid and subhumic climate is suggested. The considerable frequency of restorers and partial restorers in male fertile cytoplasm suggests that the restoration system evolved before the msm1-type cytoplasm.

In the nuclear genotype near-isogenic with either 'Adorra' or 'Ris $\phi$  1508', msm1 plants heterozygous for *Rfm1a* produced 98.6 or 98.5% of the protein content in the respective recurrent pollen parent varieties. The amino acid compositions of the derivatives differed little from those of the varieties. In the derivatives, a consistent decrease was found in tryptophan, and consistent increases in isoleucine, phenyalanine, lysine, histidine, and arginine. In relation to glucose consumption, the bioenergetic cost calculated for the amino acid patterns found in the restored msm1 derivatives was slightly higher than that for the near-isogenic pollen parent varieties. The results suggest that the restorer gene in the heterozygous state normalizes the physiology of msml cytoplasm to a great extent.

Key words: Hordeum vulgare – Cytoplasmic male sterility – Restorer gene – Polymorphism – Kernel protein and amino acids

## Introduction

Cytoplasmic male sterility in barley (Hordeum vulgare L. s.l.) was found as a natural variant in an Israeli strain of wild barley, H. vulgare ssp. spontaneum (C. Koch) Thellung (Ahokas 1979a). The original strain is a carrier of a dominant restorer gene of fertility, designated Rfm1a. No cultivated barley tested has proved to carry a restorer gene, though partial restorers have appeared (Ahokas 1979a, 1979b). It was assumed that complete restorers exist in the wild originator of barley. This has proved to be true (Ahokas 1979b).

In Israel, ssp. *spontaneum* occupies an extraordinarily wide diversity of habitats: from mesic Mediterranean to desert (Harlan and Zohary 1966; Nevo et al. 1979a, 1979b). The Israeli populations of ssp. *spontaneum* have been found to be highly variable in flavonoids (Fröst and Holm 1975), in their resistance to fungal pathogens (Fischbeck et al. 1976; Wahl et al. 1978; Segal et al. 1979; Dr. J.G. Moseman, pers. commun.), in morphology (Kamm 1977), in isozymes by starch gel electrophoresis and in morphology (Brown et al. 1978a; Nevo et al. 1979a, 1979b), in hordein patterns using two electrophoretic systems (Doll and Brown 1979), and in DBC protein content (Ahokas, unpublished). The pattern of isozyme variation has been explained by natural selection (Nevo et al. 1979a, 1979b). This evidently applies to the resistance to pathogens (Wahl et al. 1978).

The present paper describes the restoration ability of 82 accessions of ssp. *spontaneum* from Israel. The mode of inheritance of the restorer genes found in this and an earlier set (Ahokas 1979b) was studied. The protein content and amino acid composition were studied in two derivatives heterozygous for Rfm1a. An increase in protein content was ascribed to msm1 cytoplasm in unrestored derivatives (Ahokas 1979c).

#### Material and Methods

The field conditions (SE Finland, ca.  $61^{\circ}$ N) and methods of cultivation were the same as described in Ahokas (1979b), the soil being fortified each year with boron. Boron is necessary for male fertility in barley (Löhnis 1940; Simojoki 1972). The early season of 1979 was somewhat warmer (in mean  $1.3^{\circ}$ C) than that of the previous test season of 1978. The seed sets were tested by bagging at least 10 spikes on several plants at random, preferably on the earliest and next earliest stems. The mean plant density in the test bench was 60 plants per m<sup>2</sup> (Table 1). The wild barley pollen donors contain both true wild barleys and types with distinct signs of introgression from cultivated barley. Notwithstanding that, they are all called wild or *spontaneum* barleys in this paper.

For the protein determinations, the variety used as recurrent pollen parent and its near-isogenic derivative were grown in alternate rows as described in Ahokas (1979c). The derivative rows displayed a 1:1 segregation of restored and male sterile plants. The same number of the near-isogenic fertile derivatives and of the variety itself were harvested at random in the inner part of the rows. Grinding was carried out as previously (Ahokas 1979c). The protein was estimated with the Kjeldahl method. The N values were corrected for reagent control (sucrose) and for recovery of N (alanine). Tryptophan was determined by spectrophotofluorometry (Aminco Bowman) as described in Ahokas (1978a), the other amino acids by automatic analysis (Jeol 6 AH). The meals were hydrolyzed with 6N HCl at 110°C for 22 h. Elution was carried out according to Jeppsson and Karlsson (1972). The automatic analysis were purchased from the Technical Research Centre of Finland, and were performed by Drs. M. Kiesvaara and T. Hattula. All protein and amino acid values are means of two determinations.

#### Results

Of the 82 pollen parent accessions of wild barley tested in the 1979 season 31.5 (38%) were maintainers of sterility, 39 (48%) were partial restorers with a detected restoration ability ranging from 0.5 to 52.3%, and 11.5 (14%) were restorers of fertility, with a detected restoration ability ranging from 90.6 to 97.2% (Table 1). The reasons for the decrease from complete seed sets in the bagged restored  $F_1$  spikes are presented elsewhere (Ahokas and Hockett 1980). One non-restorer and another restorer plant were found in the accession PI 296850. In this paper, these pollen parent plants are designated PI 296850-11/7 and PI 296850-16/7, the former being the non-restorer and the latter the restorer. These two isolates of PI 296850 are difficult to distinguish by their morphology. The two genotypes may, however, represent a spontaneous cross in the past, and the segregation therefrom. The distribution of the accessions in different fertility classes is presented in Table 2, along with the 31 crosses tested in the season of 1978 (Ahokas 1979b). All these crosses have an 'Adorra'-type genetic background in the msm1 seed parent. Not all different seed parent backgrounds are comparable (Ahokas 1979b). Sterility due to chromosomal interchanges is negligible: 99 of these spontaneum accessions have been crossed with normal 'Adorra', and evidently only three accessions (collected in Sde Hemed) had partial  $F_1$  fertility when selfed or backcrossed, suggesting heterozygosity for the rearrangements. These three accessions were classified as maintainers of sterility.

The restorer accessions comprise both true wild or weedy specimens and a single accession with signs of introgression from cultivated, probably six-rowed barley (PI 284755), accessions with relatively short anthers (e.g. PI 296899) and rather long anthers. However, the longest anthers the author has ever seen in *H. vulgare* s.l. were found in an accession maintaining male sterility, PI 296796. These robust plants bore anthers with an anthesis length of up to 8 mm.

The original collection regions in Israel could be traced for most of the accessions, and for some the collection locality was also available (Anonymous 1968; Wahl 1972; Drs. Nava Eshed and J.G. Moseman, pers. commun.). The distribution of the accessions in different restoration classes is presented in Table 2. The sample is concentrated in the region of the Judean Foothills. The collection area in Judean Foothills, as limited by the 1948 boundary, is almost entirely covered by 'areas 1' of Dan (1977: 'thick Nari cover on chalks and marls'), or the intervening alluvial valleys. In the other ten regions, this type of lime crust occurs sporadically or not at all. Significantly more restoration ability was found in the accessions from Judean Foothills than in those from the other regions (Table 2; significance for two-tailed test, P < 0.025. In general, the restoration ability is assumed to be more common in the accessions from the central inland regions. Wild barley occurs in more southern areas in the Negev. These southernmost, strikingly arid regions should perhaps be considered a separate, major region.

Since the segregation of the winter growth habit hampers the recording of the  $F_2$  generation, backcrossed  $F_1$ generations were scored to determine the mode of inheritance of the restorer genes. The results suggest that all the restorer accessions are carriers of a single dominant restorer gene (Table 3).

The restorer accessions were screened for the presence of any *msml*-type sterilizing cytoplasm. As carriers of a H. Ahokas: Cytoplasmic Male Sterility in Barley. Part 7

 Table 1. Maintainers of sterility, restorers and partial restorers of fertility among the tested PI accessions of wild barley (ssp. spontaneum) from Israel in the season of 1979

Pollen parent	Maternal back-	Number of pollen	Number of F <sub>1</sub> plants	Period of bagging	Number of bagged	Seed set in bagged spikes		Classifi- cation <sup>b</sup>
parent	ground <sup>a</sup>	donor plants tested	i punto	(days)	spikes	Florets with seed/without seed	%	oution
PI 282572	1	1	17	11	12	19/168	10.2	S-PF
PI 282575	2	1	17	7	11	0/191	0.0	S
PI 282583	2	1	18	5	14	0/232	0.0	S
PI 282586	1	1	19	7	10	0/160	0.0	S
PI 282591	2	1	18	8	14	0/231	0.0	S
PI 282599	1	1	17	10	13	0/209	0.0	S
PI 282608	1	1	19	5	13	85/115	42.5	S-PF
PI 282609	2	1	15	5	12	97/93	51.1	S-PF
PI 282611	1,2	2	36	6	14	0/200	0.0	S
PI 282611,	1	1	16	8	12	0/154	0.0	S
atypical <sup>c</sup>								
PI 282613	2	1	14	5	11	0/186	0.0	S
PI 282616	1	1	15	15	12	1/198	0.5	S-PF
PI 282620	1	1	12	16	12	0/219	0.0	S
PI 282621	2	1	15	7	15	0/250	0.0	S
PI 282631	2	1	32	6	16	5/254	1.9	S-PF
PI 282635	1	1	24	11	24	0/391	0.0	S
PI 282636	1,3	1	21	15	18	297/ 20	93.7	Restorer
PI 282637	2	1	9	15	16	260/ 11	95.9	Restore
PI 282638	2	1	14	4	11	81/119	40.5	S-PF
PI 282640	1	1	19	9	11	11/141	7.2	S-PF
PI 282642	2	1	12	16	16	137/125	52.3	S-PF
PI 282644	2	1	15	8	13	15/174	7.9	S-PF
PI 282645	1	1	34	9	24	7/334	2.1	S-PF
PI 282646	1,3	1	29	13	19	272/ 10	96.5	Restores
PI 282663	1,5	1	17	8	11	0/172	0.0	S
PI 282665	1	1	17	4	10	34/124	21.5	S-PF
PI 282666	2	1	17	7	10	18/222	7.5	S-PF
PI 282669	1	1	16	8	13	46/188	19.7	S-PF
PI 282670	1	1	10	35	13	0/253	0.0	S
		1	17	9	14	17/211	7.5	S-PF
PI 282672	1						0.0	S
PI 282674	2	1	14	11	15 12	0/251 3/193	1.5	S S-PF
PI 282679	1	1	19 24	5		309/ 32	90.6	Restorer
PI 284742	1,3	1	24	15	21 21	309/ 32 334/ 15	95.7	Restorer
PI 284743	1,3	1	22	11		-	91.2	Restorer
PI 284753	2	1	22	12	22	332/ 32	0.0	S
PI 284754	1,2	1	31	5	12	0/169	97.2	Restore
PI 284755	2,3	1	30	15	38	590/ 17		
PI 296796	1	1	17	8	11	0/200	0.0	S
PI 296812	1	1	16	14	12	0/202	0.0	S
PI 296813	2	1	18	4	13	0/217	0.0	
PI 296815	2	1	19	9	11	1/172	0.6	S-PF
PI 296827	2	1	13	6	11	0/192	0.0	S
PI 296834	1	1	16	5	12	92/100	47.9	
PI 296838	2	1	20	15	26	406/ 17	96.0	Restore
PI 296839	1	1	17	5	11	0/185	0.0	S
PI 296850 <sup>e</sup>	1	1	13	17	12	0/192	0.0	S
PI 296850 <sup>f</sup>	2	1	2	52	10	175/ 12	93.6	Restore
PI 296853	2,3	1	24	12	29	409/22	94.9	Restore
PI 296854	1,2	2	31	3	11	0/188	0.0	S
PI 296861	1	1	15	6	11	0/172	0.0	S
PI 296863	1	2	38	7	20	3/308	1.0	S-PF
PI 296864	1	1	18	13	10	27/152	15.1	S-PF
PI 296865	2	2	25	16	15	9/227	3.8	S-PF
PI 296873	1,2	2	34	7	18	0/271	0.0	S
PI 296875	2	1	17	14	11	1/183	0.5	S-PF

1	n	2
1	7	o

Table 1. (continued)

Pollen parent	Maternal back-	Number of pollen	• Number of F, plants	Period of bagging	Number of bagged	Seed set in bagged spikes		Classifi- cation <sup>b</sup>
	ground <sup>a</sup>	donor plants tested	1 1	(days)	spikes	Florets with seed/without seed	%	
P1 296877	2	1	10	16	12	1/220	0.5	S-PF
PI 296881	2	1	16	15	12	1/185	0.5	S-PF
PI 296882	2	2	27	7	18	43/238	15.3	S-PF
PI 296884	1	1	16	11	15	44/185	19.2	S-PF
PI 296886	2	1	16	15	16	59/204	22.4	S-PF
PI 296892	1	1	22	13	14	13/242	5.1	S-PF
PI 296899	1,3	2	29	10	33	503/ 30	94.4	Restorerd
PI 296904	2	1	16	9	13	48/152	24.0	S-PF
PI 296915	2	1	20	9	14	16/220	6.8	S-PF
PI 296932	2	1	17	16	12	0/187	0.0	S
PI 296933	2	1	18	7	12	0/189	0.0	S
PI 296934	1	1	24	11	16	0/254	0.0	S
PI 296944	2,3	1	26	15	25	391/ 17	95.8	Restorer <sup>d</sup>
PI 296945	2	1	15	11	16	3/252	1.2	S-PF
PI 296952	1	1	20	7	10	66/106	38.4	S-PF
PI 296956	2	1	18	7	14	76/144	34.5	S-PF
PI 349807	2	1	17	4	10	44/121	26.7	S-PF
PI 349808	1,2	2	20	11	13	20/199	9.1	S-PF
PI 349809	1	1	18	15	10	6/154	3.8	S-PF
PI 354926	1	1	22	5	12	0/189	0.0	S
PI 354927	1	1	32	4	12	0/183	0.0	S
PI 354928	2	1	14	11	11	49/114	30.1	S-PF
PI 354929	1	1	17	16	12	4/178	2.2	S-PF
PI 354936	2	1	17	8	12	16/195	7.6	S-PF
PI 354942	2	1	33	11	17	0/256	0.0	S
PI 354944	1	1	18	10	11	1/165	0.6	S-PF
PI 354947	1	1	20	10	12	0/204	0.0	S
PI 354949	1	1	12	11	11	0/177	0.0	S
Control, msn	n1/6*Adorra	or msm1/8*Adorr	a	47	87	0/1875	0.0	S

<sup>a</sup> 1 = msm1/5\*Adorra; 2 = msm1/6\*Adorra; 3 = msm1/7\*Adorra

<sup>b</sup> S = male sterile; S-PF = male sterile to partially male fertile

<sup>c</sup> Plant organs larger than in the typical plants of the accession

<sup>d</sup> Additional six plants classified as male fertile in the greenhouse in winter 1978-1979

e Hereafter called PI 296850-11/7

f Hereafter called PI 296850-16/7

single dominant restorer gene, the  $BC_1$ - $F_1$  generation with 'Adorra' (a non-restorer) as the recurrent pollen parent will reveal the tentative presence of such a cytoplasm. No sterile segregant was found, and the hypothesis of the *msml*-type cytoplasm can be rejected for all the accessions (P < 0.001) (Table 4). It is also improbable that such a cytoplasm is present in any of the partial restorers.

Four restorer genes were tested for allelism. The test consisted of crossing a heterozygous and a homozygous restorer in *msm1* cytoplasm, and recording segregating and non-segregating  $F_2$  families (Table 5). In effect, this method cannot distinguish between linked genes and allelism, but is readily able to distinguish between non-linkage versus linkage or allelism. The restorer gene in PI 284755,

PI 296856, or PI 296919 is evidently allelic to the Rfm1a allele in Sel.77-1. Further tests on allelism can be carried out when the genes have been extracted into a spring growth habit and cultivated background.

The segregation of the restorer gene Rfm1a in highly 'Adorra'-like backgrounds was recorded in the season of 1979 (Table 6). All the results fit the monohybrid dominant mode and confirm the earlier findings (Ahokas 1979a).

Some quantitative properties of the heterozygous, restored, near-isogenic msm1 derivative of 'Adorra' were compared with those of normal 'Adorra' (Tables 7 and 8), and those of a corresponding less isogenic derivative of 'Ris $\phi$  1508' with those of 'Ris $\phi$  1508' (Table 8). These derivatives are visually indistinguishable in appearance

$r_1$ spikes with an 'Adorra'-type seed parent in msm1 cyto-	
able 2. Ranked distribution of restoration ability in the accessions of wild barley from Israel. Selfing determined in bagged F	in the season of 1978 and 1979
<b>fable 2.</b> R	plasm in th

Geobotanical region	Seed set percentage <sup>a</sup>	rcen tage <sup>a</sup>										
	0.0	0.1-1.0	1.1-5.0	5.1-10.0	10.1-20.0	20.1-30.0	30.1-40.0	40.1-50.0	50.1-60.0	60.1-90.0	5.1-10.0 10.1-20.0 20.1-30.0 30.1-40.0 40.1-50.0 50.1-60.0 60.1-90.0 90.1-100.0 Total	Total
Judean Foothills Ten regions <sup>b</sup> Region unknown and atypical accessions	17 17.5° 7	<i>ო 6 ო</i>	10 2 1	5 1	s	<i>ω ω</i> −	ю I —	2 - 1	8		9 4.5 <sup>c</sup>	59 40 14
Number of accessions 10 + 31.5 tested in the season	10 + 31.5	8 + 7	6 + 7	0 + 8	1 + 5	3 + 4	1+3	0 + 3	0 + 2	0+0	2 + 11.5	31 + 82
of 1978" + 1979 Total %	41.5 36.7	15 13.3	13 11.5	8 7.1	6 5.3	7 6.2	4 3.5	3 2.7	2 1.8	0.0	13.5 11.9	113 100

<sup>b</sup> Western, Upper and Lower Galilee, Zevulon Plain, Mt. Carmel, Jordan and Bet Shean Valley, Coastal Plain, Judean (Jerusalem) Mountains, Negev <sup>c</sup> PI 296850 is scored half to the class 0.0 and half to the class 90.1-100.0, see text <sup>d</sup> Data presented in Ahokas (1979b)

Restorer	Transmission of	Transmission of	Total:	×3	P	Suggested
accession	restoration	restoration	fer tile/male	for total,		gene
	through pollen:	through egg cells:	sterile plants	expected		symbol
	ieruie/maie sterile plants <sup>a</sup>	ierue/maie sterile plants <sup>b</sup>		1:1		
PI 282636	47/47	18/17	65/ 64	0.008	>0.90	Rfm.,e
PI 282637	49/52	35/25	84/77	0.304	>0.50	Rfm.,f
PI 282646	55/63	31/33	96/98	0.549	>0.30	Rfm, g
PI 284742	32/38	24/15	56/ 53	0.083	>0.70	Rfm,h
PI 284743	43/51	18/33	61/84	3.648	>0.05	R fm.,i
PI 284753	09/99	43/51	109/111	0.018	>0.80	Rfm,,j
PI 284755	75/62	29/28	104/90	1.010	>0.20	Rfmld
PI 296838	47/53	24/53	71/ 78	0.329	>0.50	Rfm, k
PI 296850-16/7	54/46	NT	54/46	0.640	>0.30	Rfm.,1
PI 296853	59/54	22/15	81/69	0.960	>0.30	Rfm.,m
PI 296856	54/34	40/52	94/86	0.356	>0.50	Rfm1b
PI 296899	42/39	NT	42/39	0.111	>0.70	R fm.,n
PI 296919	47/64	29/30	76/94	1.906	>0.10	Rfm1c
PI 296944	17/27	29/26	46/53	0.495	>0.30	Rfm.,o

H. Ahokas: Cytoplasmic Male Sterility in Barley. Part 7

197

Table 4. Test crosses for the tentative existence of an msml-type sterilizing cytoplasm in the restorer accessions. BC<sub>1</sub>-F<sub>1</sub> segregation of the crosses PI Accession/2\*Adorra

Restorer accession (The donor of the	$BC_1$ - $F_1$ segrega	tion <sup>a</sup>
cytoplasm in the cross)	Male fertile	Male sterile
PI 282636	96	0
PI 282637	124	0
PI 282646	96	0
PI 284742	97	0
PI 284743	144	0
PI 284753	126	0
PI 284755	87 <sup>b</sup>	0
PI 296838	94	0
PI 296850-16/7	38	0
PI 296853	74	0
PI 296856	150	0
PI 296899	109	0
PI 296919	182	0
PI 296944	199	0

<sup>a</sup> Expected segregation 1:1, if a carrier of an *msm1*-type cytoplasm and a single locus restoration of fertility. Probability for each cross P < 0.001

<sup>b</sup> Involves a single partially fertile plant

from the corresponding normal varieties. Little or no significant difference could be shown by measurements (Table 7). The mean spike fertility did not differ in this pair, though the *msm1* derivative had a significantly (P <0.05) different seed set on the two highest stems. In the nuclear genetic background of 'Adorra' and 'Risø 1508', msm1 cytoplasm plus restorer gene decreased the protein  $(N \times 6.25)$  content by 1.4 or 1.5%, consistently decreased the tryptophan content of the protein, and consistently increased the isoleucine, phenylalanine, lysine, histidine, and arginine content of the protein (Table 8). The derivatives produced a higher total basic amino acid content than the normal varieties. The increase was 0.60 and 0.34 g/16 g N in the 'Adorra' and the 'Risø 1508' derivative, respectively. The relative energy required with reference to glucose (Mitra et al. 1979) to synthesize the amino acid composition of the four barley stocks is presented in Table 8. The amino acids of the derivatives involve more energy than those of the normal varieties. A small surplus of 0.2 or 0.3% remains when the lower protein content in the derivatives is taken into account (Table 8).

Table 5. Ratios of segregating to non-segregating  $F_2$  families in crosses testing the allelism of four Rfm genes

Cross <sup>a</sup>	Number of $F_2$	families <sup>b</sup>	P (Evnostad	Tentative gene symbol (and accession of the
	Segregating	Non-segregating	(Expected 1 : 1) <sup>c</sup>	(and accession of the gene)
 Rfm,,b/+ × Rfm1a/Rfm1a	7	13	0.264	<i>Rfm1b</i> (PI 296856)
$Rfm,,c/+ \times Rfm1a/Rfm1a$	6	7	1.000	Rfm1c (PI 296919)
$Rfm, b/+ \times Rfm, c/Rfm, c$	5	7	0.774	See above
$Rfm1a/+ \times Rfm, d/Rfm, d$	12	7	0.360	<i>Rfm1d</i> (PI 284755)

<sup>a</sup> Sign + refers to the recessive allele of 'Adorra'

<sup>b</sup> Families containing less than 16 plants were excluded

<sup>c</sup> Probability determined using the binomial test (Siegel 1956)

Table 6. Segregation of the restorer gene Rfm1a in 'Adorra'-type genetic background with msm1 cytoplasm. Data from the 1979 season

Test	Countable genetic background of 'Adorra'	Plants fertile/male sterile	Expected ratio	x²	Р
Transmission through pollen, F, plants	97.7%	39/44	1:1	0.301	>0.50
Transmission through egg cells, F, plants	96.1 or 98.0%	99/75	1:1	3.310	>0.05
$F_2$ segregation	96.1%	203/66	3:1	0.031	>0.80
Heterogeneity between transmissions through the gametes				1.936	>0.10

<b>D</b> .	
Disci	ission
201044	

The two genotypes in the accession PI 296850 suggest that nonisogeny occurs within a few of the accessions and that the 1-2 plant sample per accession probably did not reveal all the variation within the accessions. One accession was observed to segregate male sterility due to a tentative male sterile gene mutant.

Among the partial restorers, there may be semidominant single gene restorers. In some instances, if not in all, partial restoration depends on more than one allele or gene (Ahokas 1979b). The fertility class 90.1-100.0%, widely separated from partial restorers (Table 2), as well as the four allelism cases found, suggest that the restorer genes are likely to be in a single locus. If the criterion for allelism (Table 5) is accepted, the central inland population, and probably some other populations as well, can be regarded as at least dimorphic, and perhaps polymorphic for the restorer gene. Since the 14 carriers of the restorer gene are not in an msm1-type cytoplasm, we are faced with three distinct groups of phenomena in nature: 1) the ecophysiology and genetics of restorer gene in fertile cytoplasm, 2) the ecophysiology and genetics of msm1 cytoplasm, and 3) the interaction of restorer gene and msm1 cytoplasm. Which one is dealt with in further discussions, may be seen in the context. The physiological effects of the partial restorer genotypes may be similar to, but milder than those of the restorer gene.

The restorer gene appears independently of the msm1 cytoplasm with considerable frequency. Furthermore, a wide range of partial restorers occurs with high frequency in these areas. These observations suggest that the restorer gene evolved before the msm1 cytoplasm, whose physiological effects are quite well normalized by the gene Rfm1a, at least in the heterozygous state (Tables 7 and 8; Ahokas 1978b, 1980).

A striking part of the genetic variation found in 28 populations of wild barley in Israel (from a wider area than the present material) was correlated with some environmental factors, chiefly with combinations of temperature and humidity, and also with soil types and vegetation (Nevo et al. 1979b). It may be noted that these factors are interrelated. Nevo et al. (1979b) found the highest polymorphism in the regions with an annual rainfall of 200-600 mm in the curvilinear association. The pattern of variation was concluded to suggest the operation of natural selection in *spontaneum* barley in Israel.

Unfortunately, the accurate collection locality is known only for part of the present accessions, and no direct information about the soil type in the habitats is available. On the other hand, changes in the accessions since the collection have been assessed as small: outcrossing is minimal in the greenhouse maintenance of the stocks (Dr. J.G. Moseman, pers. commun.). Five atypical

Table 7. Effects of the restorer gene in msml cytoplasm with an 'Adorra'-type genetic background. Sample sizes 20 plants

Cytoplasm	Rfm gene <sup>a</sup>		Plant	Plant height	Spikes per plant	Total number	Seed set ]	Seed set percentage		Mean	Yield per plant	Protein %
		nuclear genotype of cv. 'Adorra'	density per m <sup>2</sup>			ou noters per spike	On two highest tillers	On the other tillers	On all tillers together	weight on total sample (mg)	Ŕ	
Adorra msm l	+ / + 100.0% Rfm1a/ + 98.8%	100.0% 98.8%	135 135	91.1 ± 0.80 92.3 ± 1.05	$4.45 \pm 0.35 \\4.55 \pm 0.33$	$17.89 \pm 0.47$ $17.73 \pm 0.43$	98.6% 97.0%	91.7% 94.1%	95.3% 95.6%	38.3 39.5	2.90 ± 0.21 3.04 ± 0.24	7.61 7.50
t (x <sup>2</sup> ) P		I		0.945 >0.30	0.209 >0.80	0.256 >0.70	(4.010) <0.05	(3.065) >0.05	(0.056) >0.80	(4.010)         (3.065)         (0.056)         Not deter-         0.446           <0.05	0.446 >0.60	Not deter- mined

Sign + refers to the recessive allele of 'Adorra'

æ

Amino acids	Variety and <i>msm1</i> derivative (countable nuclear isogeny in parenthesis)			
	'Adorra'		'Risø 1508'	
	In msm1, Rfm1a/+ (98.8%)	Normal 'Adorra', +/+ (100.0%)	In <i>msm1</i> , <i>Rfm1a</i> /+ (87.5%)	Normal 'Risø 1508', +/+ (100.0%)
Tryptophan	0.82	0.86	0.69	0.76
Aspartic acid	6.15	6.09	7.38	7.36
Threonine	3.66	3.59	4.12	4.11
Serine	4.23	4.22	4.51	4.45
Glutamic acid	19.77	19.36	14.41	14.71
Proline	9.57	9.63	6.13	5.92
Glycine	4.01	4.05	5.09	5.02
Alanine	4.31	4.28	4.99	4.91
Cysteine	1.51	1.37	1.31	1.37
Valine	4.84	4.86	5.06	5.09
Methionine	1.05	1.19	1.17	1.17
Isoleucine	3.60	3.21	3.16	2.42
Leucine	6.24	6.19	5.92	5.94
Tyrosine	2.97	3.14	2.98	2.97
Phenylalanine	4.67	4.55	3.84	3.79
Lysine	3.64	3.47	5.11	4.85
Histidine	2.22	2.18	2.60	2.56
Arginine	5.19	4.79	6.26	6.23
Protein N × 6.25	7.50	7.61	8.67	8.80
(Relative)	(98.6)	(100.0)	(98.5)	(100.0)
Relative glucose	12057	11845	11698	11501
requirement for the amino acids <sup>a</sup>	(101.8)	(100.0)	(101.7)	(100.0)
Relative glucose requirement for the amino acids per unit of meal	100.3	100.0	100.2	100.0
Spike fertility	95.6%	95.3%	96.3%	97.2%
Number of plants in sample	20	20	15	15
Plant density in the plot, plants per m <sup>2</sup>	135	135	84	84

Table 8. Amino acids in kernel protein (g/16 g N) and quantitative comparisons between heterozygous, restored *msm1* derivatives of 'Adorra' and 'Ris $\phi$  1508' and the original varieties

<sup>a</sup> Based on calculations by Mitra et al. (1979)

constant plants separated from 110 accessions probably removed the rest of the material from seed mixings. The selection since collection is low because wild barley is principally homozygous (Brown et al. 1978b). There is no conscious selection with respect to restoration among the sample.

In the present material, there is no apparent association between the distribution of restorers or partial restorers and the two climatic region systems, according to either Köppen or Thornthwaite (Rosenan and Mané 1970). To determine whether or not salt plays a role in the distribution of restoration ability, further material must be studied. Tests on material with pollen parents from two regions of Negev rather reject the association between restoration and salty habitats. These  $F_1$  plants in *msm1* cytoplasm have so far been tested only in the greenhouse environment.

The dominant restorer gene is evidently long-term in

activity, i.e. constitutive or nearly constitutive, and affects many plant organs, or even the whole plant (Ahokas 1978b, 1980). This combined with its dominance, makes the restorer gene well subjected to natural selection. A constitutive gene is expected to be correlated with a selective factor(s), which is likely to operate for long periods or even for the entire life of the plant. Many soil factors are persistent or change little during an annual's life. Evidence for the involvement of soil has been received from two unpublished observations. Partial restoration responds to the level of soil nitrogen. KNO<sub>3</sub> fertilizer, which is the beneficial ion pair for nitrate uptake in barley (Blevins et al. 1978), was found to raise partial restoration up to a certain level of fertilization, Secondly, there was a decrease in partial restoration due to a change in the soil type (from a mixture of clay, sandy loam, and Finnpeat B2 to Finnpeat B2 alone) in the greenhouse when other conditions were kept constant.

One restorer accession from Judean Foothills has recently been found in addition to those described here. Thus, 15 out of 16 restorer plants originate from hilly or mountainous regions. Such regions with slopes, valleys and top plateaux offer a great variety of soil environments. The catenary soil relationships on a hill and the adjoining valley in Judean Foothills were accurately described by Dan et al. (1972). These soils have pH values from 7.1 to 8.1, and may be highly calcerous. Clay and silt are the major particle fractions (Dan et al. 1972). 10-15 out of 16 restorer plants originate from the eroded areas 'characterized by thick Nari cover on chalks and marls' (Dan 1977: 'areas 1') or the intervening valleys. Similar marly slopes and alluvial valley soils also appear in En Kerem (for PI 296850) and in Moza (for PI 296919) in the environs of Jerusalem (Arkin 1973). The collection site for PI 296856 is not unambiguous (Anonymous 1968), but this accession is likely to originate from 'areas 1' between Beer Sheva and Arad. Therefore, the number is likely 13 or more out of 16 restorers. Nari areas are found in the inland on soft chalky and marly parent material, where the climate is subhumic or semiarid down to the 200 mm isohyet. The 'areas 1' also appear sporadically in Galilee (Dan 1977), from where one restorer besides Sel.77-1 originates.

Most, if not all, of the restorer accessions originate from the wide areas between the 200-600 mm isohyets of annual rainfall (Rosenan 1970). The highest genetic polymorphism is expected to occur under these precipitation conditions (Nevo et al. 1979b). Most, if not all, of the restorers also appear inside the areas with 30-50 rainy days per year (Rosenan 1970). It is plausible that the semiarid conditions reinforce the hypothetical effective soil factor or participate in the soil formation, e.g. affecting its carbonate content (Gal et al. 1974).

The performance of the heterozygotes for the restorer

gene is of interest from the standpoint of the possible use of this system in hybrid barley production. An increase in protein was attributed to the unrestored msml cytoplasm (Ahokas 1979c). In this cytoplasm, the heterozygotes for Rfmla have a protein content and amino acid composition very similar to that of the normal varieties, 'Adorra' and 'Risø 1508' (Tables 7 and 8). Thus, in the heterozygous state, the restorer gene to a great extent also normalizes the kernel composition, as it does many other features in the plant (Table 7; Ahokas 1978b, 1980). The small differences in spike fertility are expected to have negligible effect on the protein content (see Ahokas 1979c). Interestingly, the small surplus in the calculated glucose consumption for the amino acids was of the same magnitude in both the derivatives, though the varieties have a widely different amino acid composition and a different protein content. This suggests that the surplus is probably not due to a chance, and that the bioenergetic values determined by Mitra et al. (1979) are close to the real situation in barley. This observation of the bioenergetic contents raises the question of the slight superiority of Rfm1a heterozygotes in normal cytoplasm.

#### Acknowledgements

I am grateful for information to Nava Eshed, J.G. Moseman, Dolly Piper, and the staff of the Embassay of Israel in Helsinki. The seeds of the PI (Plant Introduction) accessions were kindly supplied by Dr. J.G. Moseman (PGGI, USDA, BARC, Beltsville, MD, USA). Some of the analyses on kernel composition were carried out with the facilities of Departments of Limnology and Botany of the University of Helsinki. The work was performed under the auspices of the Research Council for Agriculture and Forestry (the Academy of Finland), which also provided financial support.

#### Literature

- Ahokas, H. (1978a): A simple and rapid screening method for the determination of protein and tryptophan in kernel halves and small samples of barley meal. J. Sci. Food Agric. 29, 47-52
- Ahokas, H. (1978b): Cytoplasmic male sterility in barley. 2. Physiology and anther cytology of msm1. Hereditas 89, 7-21
- Ahokas, H. (1979a): Cytoplasmic male sterility in barley. Acta Agric. Scand. 29, 219-224
- Ahokas, H. (1979b): Cytoplasmic male sterility in barley. 3. Maintenance of sterility and restoration of fertility in the *msm1* cytoplasm. Euphytica 28, 409-419
- Ahokas, H. (1979c): Cytoplasmic male sterility in barley. 4. Effects of *msm1* cytoplasm and partial fertility on kernel protein and lysine. Theor. Appl. Genet. 55, 269-272
- Ahokas, H. (1980): Cytoplasmic male sterility in barley. 5. Physiological characterization of the msm 1 - Rfm 1a system. Physiol. Plant. 48, 231-238
- Ahokas, H.; Hockett, E.A. (1980): Cytoplasmic male sterility in barley. 6. Tests on performance at two different latitudes
- Anonymous (1968): Hordeum spontaneum C. Koch (PI 296777 to 296956). In: Plant inventory No. 172 (ed. Hyland, H.L.), pp. 82-86, Washington D.C.: USDA

- Arkin, Y. (1973): Map 1: 3, Geology. In: Atlas of Jerusalem (eds. Amiran, D.H.K.; Shachar, A.; Kimhi, I.; Karmon, M.; Bandel, P.). Berlin, New York: de Gruyter
- Blevins, D.G.; Hiatt, A.J.; Lowe, R.H.; Leggett, J.E. (1978): Influence of K on the uptake, translocation, and reduction of nitrate by barley seedlings. Agron. J. 70, 393-396
- Brown, A.H.D.; Nevo, E.; Zohary, D.; Dagan, O. (1978a): Genetic variation in natural populations of wild barley (*Hordeum spon*taneum). Genetica 49, 97-108
- Brown, A.H.D.; Zohary, D.; Nevo, E. (1978b): Outcrossing rates and heterozygosity in natural populations of *Hordeum spontaneum* Koch in Israel. Heredity **41**, 49-62
- Dan, J. (1977): The distribution and origin of Nari and other lime crusts in Israel. Isr. J. Earth-Sci. 26, 68-83
- Dan, J.; Yaalon, D.H.; Koyumdjisky, H. (1972): Catenary soil relationships in Israel. 2. The Bet Guvrin catena on chalk and Nari limestone crust in the Shefela. Isr. J. Earth-Sci. 21, 99-114
- Doll, H.; Brown, A.H.D. (1979): Hordein variation in wild (Hordeum spontaneum) and cultivated (H. vulgare) barley. Canad. J. Genet. Cytol. 21, 391-404
- Fischbeck, G.; Schwarzbach, E.; Sobel, Z.; Wahl, I. (1976): Mehltauresistenz aus israelischen Populationen der zweizeiligen Wildgerste (Hordeum spontaneum). Z. Pflanzenzücht. 76, 163-166
- Fröst, S.; Holm, G. (1975): Variation of flavonoid patterns in Hordeum spontaneum and H. agriocrithon. Hereditas 80, 167-172
- Gal, M.; Amiel, A.J.; Ravikovitch, S. (1974): Clay mineral distribution and origin in the soil types of Israel. J. Soil Sci. 25, 79-89
- Harlan, J.R.; Zohary, D. (1966): Distribution of wild wheats and barley. Science 153, 1074-1080
- Jeppsson, J.O.; Karlsson, I.M. (1972): Ion-exchange chromatography of physiological sulphur amino acids on a highly crosslinked resin. J. Chromatogr. 72, 93-103
- Kamm, A. (1977): The range of brittle types of Cerealia barleys in Israel. Pamphlet 165, pp. 43 Bet Dagan: Agricult. Res. Organization
- Löhnis, M.P. (1940): Histology of boron deficiency in plants. Meded. Landbouwhoogeschool Wageningen 44(3), 1-36
- Mitra, R.K.; Bhatia, C.R.; Rabson, R. (1979): Bioenergetic cost of altering the amino acid composition of cereal grains. Cereal Chem. 56, 249-252

- Nevo, E.; Brown, A.H.D.; Zohary, D. (1979a): Genetic diversity in the wild progenitor of barley in Israel. Experientia 35, 1027-1029
- Nevo, E.; Zohary, D.; Brown, A.H.D.; Haber, M. (1979b): Genetic diversity and environmental associations of wild barley, *Hordeum spontaneum*, in Israel. Evolution 33, 815-833
- Rosenan, N. (1970): Rainfall. In: Atlas of Israel (eds. Amiran, D.H.K.; Elster, J.; Gilead, M.; Rosenan, N.; Kadmon, N.; Paran, U.). Jerusalem: Ministry of Labour and Amsterdam: Elsevier
- Rosenan, N.; Mané, U. (1970): Climatic regions, radiation, evaporation, wind, sharav. In: Atlas of Israel (eds.: Amiran, D.H.K.; Elster, J.; Gilead, M.; Rosenan, N.; Kadmon, N.; Paran, U.). Jerusalem: Ministry of Labour and Amsterdam: Elsevier
- Segal, A.; Fischbeck, G.; Wahl, I. (1979): Types of protection of wild barley, *Hordeum spontaneum* C. Koch, against *Erysiphe* graminis hordei in natural populations in Israel. Phytoparasitica 7, 49
- Siegel, S. (1956): Nonparametric statistics for the behavioral sciences. Tokyo: Kogakusha
- Simojoki, P. (1972): Tuloksia ochran boorilannoituskokeista. Ann. Agric. Fenn. 11, 333-341 (Finnish with English abstract)
- Wahl, I. (1972): Hordeum (hybrid) (PI 349803 to 349838). In:
   Plant Inventory No. 178 (ed.: Hyland, H.L.). pp. 90-91,
   Washington D.C.: USDA
- Wahl, I.; Eshed, N.; Segal, A.; Sobel, Z. (1978): Significance of wild relatives of small grains and other wild grasses in cereal powdery mildews. In: The Powdery Mildews (ed.: Spencer, D.M.). pp. 83-100. London, New York, San Francisco: Academic Press

Received January 25, 1980 Communicated by D. von Wettstein

Mr. H. Ahokas Department of Genetics University of Helsinki P. Rautatiekatu 13 00100 Helsinki 10 (Finland)